Research Article

DOI: 10.5455/2320-6012.ijrms20141142

The cytokine gene polymorphisms (TNF- α , IL-10 And IFN- γ) and the role of inflammatory cytokines in diabetic neuropathy

M. Ramesh*, Konathala Geetha Kumari, G. Sudhakar

Department of Human Genetics, Andhra University, Visakhapatnam - 530003, Andhra Pradesh, India

Received: 22 August 2014 Accepted: 6 September 2014

*Correspondence:

Dr. M. Ramesh,

E-mail: ramesh_mandarapu@rediffmail.com

Copyright: © the author(s), publisher and licensee Medip Academy. This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License, which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

ABSTRACT

Background: One of the most frequently-occurring micro vascular complications is diabetic neuropathy (DN). Diabetic nephropathy (DN) affects approximately one third of people with type 1 or type 2 diabetes mellitus The objective of the study is an attempt to examine functional SNPs primarily at the position on gene of TNF- α (-308 G/A, rs 1800629), IL-10 (-1082 G/A, rs 1800896) and IFN γ (+874 A/T, rs 62559044) in order to establish their association with peripheral neuropathy patients with type 2 diabetes.

Methods: 150 cases presenting Diabetic neuropathy and 160 cases of age and sex matched healthy controls were included in the study. ARMS PCR was done for genotyping of TNF- α (-308), IL-10 (1082 G/A) and IFN γ (+874) polymorphism using allele specific primers for detection of single nucleotide polymorphisms. Analysis of the data was carried out using Epi Info 5 software. In addition, the gene frequencies were estimated and goodness of fit between the observed and expected phenotype frequencies was tested. Multifactor Dimensionality Reduction (MDR) analysis was performed to study case-control data and gene-gene interactions.

Results: The results revealed that the chi- square test for heterogeneity for IL-10 system was found to be significant ($\chi^2 = 16.2380$; d.f = 2; p >0.001) between patients and controls, indicating a significant departure from the HWE. Thus, the test of association of both homogeneity and heterogeneity of IL-10 showed a significant difference, indicating an association of IL-10 with diabetic neuropathy.SNPs at position -308 promoter gene of TNF- α and IFN γ (+874) were not significantly associated with development of Diabetic Neuropathy.

Conclusion: This case-control study suggests that IL-10-1082G/G polymorphism is associated with the susceptibility to diabetic neuropathy in type 2 DM patients. IL-10 serves as an important bio marker in Indian population for their susceptibility to Diabetic Neuropathy as it may play a role in alteration of IL-10 production and the inflammatory responses.

Keywords: Diabetic Neuropathy, Tumor Necrosis Factor Alpha, Interleukin – 10, Interferon Gamma

INTRODUCTION

Diabetes mellitus (DM) is one of the most widespread chronic diseases in the world. It can be caused by the genetically predisposed lack of insulin or by the body unresponsiveness to insulin resulting in elevated blood sugar levels. According to the International Diabetes Federation, diabetes affects more than 230 million people worldwide and is expected to affect 350 million by 2025. In 2003, the five countries with the largest number of

people with diabetes were India (35.5 million), China (23.8 million), the United States (16 million), Russia (9.7 million) and Japan (6.7 million). Untreated diabetes may cause severe health complications which can be largely divided into macro vascular and micro vascular complications. The macro vascular complications include cerebrovascular disease, coronary heart disease, and peripheral vascular disease. The micro vascular complications include diabetic retinopathy, diabetic neuropathy, and diabetic nephropathy.

Cytokines are key mediators which regulate immune response; and their expression by immune cells depends on several factors such as infection, inflammation, hormonal conditions and also relevant gene polymorphisms.² Recent studies have shown that inflammation, and more specifically pro-inflammatory cytokines, play determinant role in the development of micro vascular diabetic complications. One of the most frequently occurring micro vascular complications is diabetic neuropathy (DN). Diabetic nephropathy (DN) affects approximately one third of people with type 1 or type 2 diabetes mellitus.³ It is a multifactorial disease. It develops as a result of hyperglycemia-induced local metabolic, enzymatic and micro vascular changes. The disease gradually progresses and involves small and large sensory fibers. Diabetic neuropathy is a decrease in nerve function typically affecting the lower limbs in people with diabetes. The peripheral nerves become damaged by persistently elevated blood sugar levels. These results in significant disability and morbidity.⁵ Complications of diabetic neuropathy include severe pain, loss of ambulation and increased risk of foot ulceration and amputation. Lifetime risk of foot amputation is 15% in patients with diabetic neuropathy. Different hypotheses have been proposed to explain the various modes of progression of diabetic neuropathy. It has been suggested that consumption of oral hypoglycemic agents such as glyburide⁷ and angiotensin converting enzyme inhibitors (ACE) inhibit the progression of neuropathy irrespective of blood glucose level.8 Early diagnosis and treatment of diabetic neuropathy is important for preventing secondary complications and improving quality of life. Considering that diabetes affects an estimated 177 million people worldwide, more than 20 million people suffer from diabetic neuropathy with a remarkable range of clinical manifestations.

In recent years, our knowledge of the pathophysiological processes that lead to diabetic nephropathy has notably improved on a genetic and molecular level.

Most human diseases linked to specific genetic polymorphisms are chronic in nature. Among the few known genetic polymorphisms that seem to affect the risk progression of infection, single-nucleotide polymorphisms in the cytokine cascade stand out. 10 TNFα is a multifunctional cytokine. It can directly inhibit phosphorylation of insulin receptor's substrate and reduce glucose uptake by peripheral tissues. 11 In human, the gene is located on chromosome # 6 [p21.3]. ¹² Several studies have shown that SNP at position -308 A/G were associated with various inflammatory conditions. 13 Interleukin [IL-10] is a potent inflammatory cytokine. The main function of IL-10 is terminating the inflammatory signal in inflammatory cells. It promotes B cell activation. The human IL-10 gene is located on chromosome # 1 [1q31-q32] and encodes for five exons. It has been shown to limit the cascade of proinflammatory cytokines activation¹⁴ and to down regulate T cell-mediated immune responses. 15 INF-y is

aTh1 cytokine which supports the immune system to perform cytolysis of target cells and also was reported to be increased in diabetes mellitus. ¹⁶ IFN- γ gene intron-1 polymorphism was speculated to influence immune complex disease susceptibility which is characterized by an imbalance of various immunoregulatory systems. ¹⁷ The gene is located on chromosome # 12 [12q14].

The objectives of the study is an attempt to examine functional SNPs primarily at the position on gene of TNF- α (-308 G/A, rs 1800629), IL-10 (-1082 G/A, rs 1800896) and IFN γ (+874 A/T, rs 62559044) in order to establish their association with peripheral neuropathy patients with type 2 diabetes.

METHODS

A total of 150 patients (85 males and 65 females) presenting diabetic neuropathy attending local Government King George General Hospital, Visakhapatnam, Andhra Pradesh were included in the study. The diagnosis of diabetic neuropathy was established by clinical analysis. 160 members of age and sex matched healthy individuals with no known history of any disease were taken as controls (85 males and 75 females). All the subjects were examined clinically and information pertaining to age, sex, habits and health status were recorded. The patient's ages were ranged between 30 and 80 years. Blood samples were collected in sterile vials containing 15% EDTA as an anticoagulant from both controls and patients for DNA isolation. DNA was isolated by salting out method. 18 All the three cytokine gene polymorphisms were typed by using amplification refractory mutation system polymerase chain reaction [ARMS – PCR] 19 was done for genotyping of TNF- α (-308 G/A), IL-10 (-1082 G/A) and IFN γ (+874 A/T) polymorphism using allele specific primers for detection of single nucleotide polymorphisms. The amplified products were separated on 2% agarose gels stained with ethidium bromide and visualized under a UV transilluminator.

Analysis of the data was carried out using Epi Info 5 software. In addition, the gene frequencies were estimated by using maximum likelihood methods²⁰ and goodness of fit between the observed and expected tested.²¹ phenotype frequencies were Genotype frequencies were checked for deviation from Hardy-Weinberg equilibrium and were not significantly different from those predicted. Odds ratios and 95% confidence interval (95% CI) were calculated to assess the strength of the relationship between the IL-10 polymorphisms with diabetic neuropathy. Pooled odds ratios and relative risk were calculated by the randomeffects method.²² For odds ratio, confidence interval was calculated. Increased risk was calculated using the formula: Increased $5Risk = (Relative Risk - 1.00) \times 100$. The significance level was 5%.

Multifactor Dimensionality Reduction (MDR) analysis was performed using MDR software (v. 3.0.2) to study

case-control data, gene-gene interactions, and gene-environment interactions. ^{23,24} Best models with possible combinations of the polymorphisms were considered based on 10-fold cross validation and maximum testing accuracy. Once MDR identifies the best combination of factors, the final step is to determine which multifactor levels (genotypes) are high risk and which are at low risk using the entire data set. This final evaluation is conducted with a threshold ratio that is determined by the ratio of the number of affected individuals divided by the number of unaffected individuals in the data.

RESULTS

Distribution of TNF- α , IL-10 and IFN γ polymorphisms in the study groups is presented in Table 1. The genotyping was done for the entire group of 150 patients with diabetic neuropathy and 160 controls. The genotypic distributions of TNF- α A/A, A/G and G/G genotypes in

diabetic neuropathy patients were 17.3%, 56% and 26.7% respectively and in controls it was 17.5%, 53.7% and 28.7% respectively. The genotypic distributions of IFNγ A/A, A/T and T/T genotypes were 30%, 48.7% and 21.3% in diabetic neuropathy patients and in controls it was 21.9%, 53.1% and 25% respectively. According to Hardy-Weinberg equilibrium model, the frequencies of observed genotypes of both patients and controls did not deviate significantly from those expected (TNF- α : χ^2 = 2.5474; df = 1, p>0.10; IFN γ : χ^2 = 0.0594; df = 1, p>0.80). Observed genotype frequencies of IL-10 A/A, G/A and G/G phenotypes in patients was 18%, 23.3% and 58.7% and in controls, it was 8.1%, 43.1% and 48.7% respectively. Our results indicate that there is a significant difference in the genotype frequencies in the IL-10 gene at this position i.e. -1082 amongst the diabetic neuropathy patients ($\chi^2 = 29.3594$; d.f = 1; p > 0.001) indicating a significant deviation from the Hardy-Weinberg equilibrium.

Table 1: Distribution of the TNF-α, IL-10 and IFNγ phenotypes in patients and controls.

System Cytokine	Phenotype	Diabetic (Patients	Neuropathy s)	Controls		
		Obs.	Exp.	Obs.	Exp.	
	A/A	26	30.38	28	30.98	
	A/G	84	74.25	86	78.85	
TNF-α	G/G	40	45.37	46	50.18	
(-308 G/A)	Total	150	150.00	160	160.00	
		$\chi^2 = 2$.5474	$\chi^2 = 1.2831$		
		(0.20>P>0.10)		(0.30>P>0.20)		
	G/G	27	13.05	13	13.92	
	G/A	35	62.39	69	66.55	
IL-10	A/A	88	74.56	78	79.53	
(-1082 G/A)	Total	150	150.00	160	160.00	
		$\chi^2 = 29$	0.3594	$\chi^2 = 0.1804$		
		(P>0.	001)	(0.70>P>0.50)		
		45	44.56	35	37.64	
IFNγ+874	A/A	73	74.39	85	79.93	
	A/T	32	31.05	40	42.43	
	T/T	150	150.00	160	160.00	
	Total	$\chi^2 = 0.0$	0594	$\chi^2 = 0.6460$		
		(0.90>P>	>0.80)	(0.50>P>0.30)		

The allelic distributions of TNF- α , IL-10 and IFN γ polymorphisms are given in Table 2. The frequency of the A and G alleles in diabetic neuropathy patients were 45% and 55% and in controls it was 44% and 56% for TNF- α . For IL-10, the frequency of G and T alleles were 70% and 29% in patients. Same frequencies were observed in controls also. The frequency of A and T alleles in patients were 54% and 45% and in controls were 48% and 51% for IFN γ . The results revealed that the chi- square test for heterogeneity for IL-10 system was found to be significant ($\chi^2 = 16.2380$; d.f = 2; p >0.001) between patients and controls, indicating a significant departure from the HWE. Thus, the test of association of both homogeneity and heterogeneity of IL-

10 showed a significant difference, indicating an association of IL-10 with diabetic neuropathy.

Association between different combinatory forms of alleles was estimated. Test of association of TNF- α , IL-10 and IFN γ phenotypes with the disease condition compared to the control group, the odds ratio and relative risks for each genotype versus the other two are shown in Table 3. In TNF- α , the heterozygous (GA) individuals are equally likely to get the disease (RR = 1.04), with an overall odds ratio of 1.10 (95% CI: 0.68, 1.76; p = 0.6907). Individuals with the other two groups (AA and GG) were at a reduced risk of diabetic neuropathy. In IL-10, the Risk estimates show a significant association of

GG and AA phenotypes with diabetic neuropathy individuals (RR = 2.22 & 1.20 respectively), with an overall odds ratio of 2.48 (95% CI: 1.17, 5.33, p = 0.0095) and 1.49 (95% CI: 0.93, 0.240, p = 0.0802) respectively. The result shows an increased risk of 100% and 0.49% more, indicating that individuals with GG phenotypes are 2 times more likely to get the disease when compared with the other phenotypes of the IL-10.

In IFN γ the Risk estimates show a significant association of AA phenotypes with diabetic neuropathy individuals (RR = 1.37), with an overall odds ratio of 1.53 (95% CI: 0.89, 2.64, p = 0.1022). The result shows an increased risk of 30% more, indicating that individuals with AA phenotype are 1.3 times more likely to get the disease when compared with the other phenotypes of the IFN γ .

Table 2: Allele frequencies in diabetic neuropathy patients and controls.

System (Allele)	Diabetic Neuropathy	Controls	Intergroup Heterogeneity	d.f
TNF-α A G	$\begin{array}{c} 0.4500 \pm 0.0287 \\ 0.5500 \pm 0.0287 \end{array}$	$\begin{array}{c} 0.4400 \pm 0.0277 \\ 0.5600 \pm 0.0277 \end{array}$	0.1932 (0.95>p>0.90)	2
IL-10 G A	0.2950 ± 0.0263 0.7050 ± 0.0263	$\begin{array}{c} 0.2950 \pm 0.0255 \\ 0.7050 \pm 0.0255 \end{array}$	16.2380 (p>0.001)	2
IFN γ A T	$\begin{array}{c} 0.5450 \pm 0.0287 \\ 0.4550 \pm 0.0287 \end{array}$	$\begin{array}{c} 0.4850 \pm 0.0279 \\ 0.5150 \pm 0.0279 \end{array}$	2.7305 (0.30>P>0.20)	2

Table 3: Test of Association, Relative Risk, Odds Ratio and 95% Confidence Interval Estimates of TNF α , IL-10 and IFN γ Phenotypes in DN patients and Control Group.

System	Phenotype	Control	Diabetic Neuropathy						
	combinations	(n)	(n)	RR	OR	95% CI	χ^2 values	p-value	
TNF α	AA vs GG + GA	28	26	0.99	0.99	0.53-1.85	0.00	0.9691	
	GA vs GG + AA	86	84	1.04	1.10	0.68-1.76	0.16	0.6907	
	GG vs GA + AA	46	40	0.93	0.90	0.53-1.53	0.17	0.6822	
	GG vs GA + AA	13	27	2.22	2.48	1.17-5.33	6.72	0.0095	
IL-10	GA vs GG + AA	69	35	0.54	0.40	0.24-0.67	13.60	0.0002	
	AA vs GG + GA	78	88	1.20	1.49	0.93-2.40	3.06	0.0802	
IFN γ	AA vs TT + TA	35	45	1.37	1.53	0.89-2.64	2.67	0.1022	
	TA vs TT + AA	85	73	0.92	0.84	0.52-1.34	0.62	0.4326	
	$TT \ vs \ TA + AA$	40	32	0.85	0.81	0.46-1.43	0.58	0.4448	

Table 4: Results of MDR analysis on genetic factors.

No. of loci	Polymorphism Model	Testing Accuracy	CVC	Prediction Error (%)
1	IL-10	0.5990	10/10	40.10*
2	IL-10, IFN-γ	0.5612	7/10	43.88
3	IL-10, IFN-γ, TNF-α	0.5946	10/10	40.54*

^{*}P\le 0.01 based on 1000 permutations.

MDR software was used to analyze the interaction of the 3 factors that may affect the diabetic neuropathy, and the results were detailed in Tables 4 and 5. We found that the cross-validation (CV) consistency of the three-factor model (IL-10*IFN- γ *TNF- α) was maximal (10/10), and the accuracy of the test samples was the highest (0.5946). Permutation testing was used to perform a hypothesis test and evaluate its statistical significance. Thus, the three-factor interaction model was the best model, which shows

that there was an interaction between the three SNP's (p \leq 0.01). Table 5 summarizes the three locus genotype combinations associated with high risk and with low risk, along with the corresponding distribution of cases and of controls, for each multilocus genotype combination. The cell is labeled as either high risk if the case—control ratio reaches or exceeds a predetermined threshold (for example, \geq 1) and low risk if it does not reach this threshold (Fig 1). Figure 2 illustrates the MDR

interaction information analysis of the three polymorphisms, represented in the form of a dendrogram. The interaction information analysis revealed a strong

synergism between the three SNPs markers TNF- α , IL-10 and IFN γ contributing to Diabetic Neuropathy.

Table 5: Distribution of high-risk and low-risk genotypes in the best three locus model.

Pattern	Multilocus-genotype combinations			No. of cases	No. of	Association
rattern	TNF- α	IL – 10	IFN – γ	(DN)	controls	with DN
1	A/A	A/A	T/T	5	2	High-risk
2	A/A	A/A	A/A	7	5	High-risk
3	A/A	G/G	T/T	2	0	High-risk
4	A/A	G/G	A/A	1	0	High-risk
5	A/A	G/G	A/T	4	1	High-risk
6	A/G	A/A	A/A	14	8	High-risk
7	A/G	A/A	A/T	26	18	High-risk
8	A/G	G/G	T/T	2	2	High-risk
9	A/G	G/G	A/T	10	3	High-risk
10	A/G	A/G	A/A	8	5	High-risk
11	G/G	A/A	T/T	6	6	High-risk
12	G/G	A/A	A/A	8	6	High-risk
13	G/G	G/G	A/A	3	0	High-risk
14	G/G	G/G	A/T	4	3	High-risk
15	A/A	A/A	A/T	5	7	Low-risk
16	A/A	A/G	T/T	0	4	Low-risk
17	A/A	A/G	A/A	1	3	Low-risk
18	A/A	A/G	A/T	1	6	Low-risk
19	A/G	A/A	T/T	10	16	Low-risk
20	A/G	G/G	A/A	0	2	Low-risk
21	A/G	A/G	T/T	5	6	Low-risk
22	A/G	A/G	A/T	9	26	Low-risk
23	G/G	A/A	A/T	7	10	Low-risk
24	G/G	G/G	T/T	1	2	Low-risk
25	G/G	A/G	T/T	1	2	Low-risk
26	G/G	A/G	A/A	3	6	Low-risk
27	G/G	A/G	A/T	7	11	Low-risk

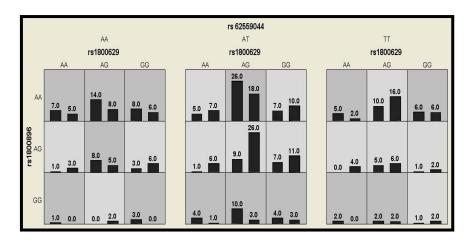


Figure 1: An MDR Analysis of the Three-factors (IL-10*IFN- γ *TNF- α) - Interaction Model of Diabetic Neuropathy.

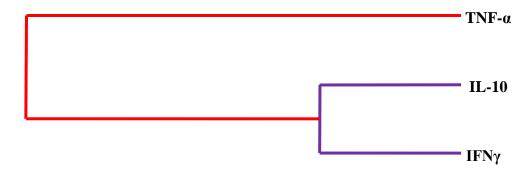


Figure 2: A Tree Diagram of the Interactions among three Factors (IL-10*IFN- γ *TNF- α) for Diabetic Neuropathy, as Analyzed by MDR.

In the cell in the figure, the left bands represent the disease case, and the right bands represent the control case. High-risk combinations are depicted as darkly shaded cells, low-risk combinations as lightly shaded cells.

DISCUSSION

The genetic basis of multifactoral diseases like diabetes probably consists of several predisposing risk factors that can interact with environmental factors to produce the disease phenotype. Diabetic neuropathy is one of the common complications of diabetes mellitus with high morbidity and impairment of quality of life reported an overall 28% of prevalence of peripheral neuropathy. 25 In diabetic patients the impaired insulin signaling and selective destruction of insulin producing beta cells in which cytokines play an important role. Certain proinflammatory cytokines are capable of interfering with insulin sensitive glucose uptake and can induce insulin resistance.²⁶ Higher levels of pro-inflammatory cytokines correlate with the incidence of neuropathy. 27 Spinal proinflammatory cytokines such as TNF-α, IFNγ and IL-10 are powerful pain-enhancing signals that contribute to neuropathic pain. 28,29 The present study focused on gene polymorphisms of pro and anti-inflammatory cytokines that may be responsible for nerve damage in diabetic neuropathy. The present study focused on three common functional SNPs primarily at the positions on genes of tumor necrosis alpha TNF-a 308G/A, interferon gamma (IFNγ) +874A/T and interleukin (IL-10) -1082G/A in order to establish their association with peripheral neuropathy in type 2 diabetes.

Inflammation is a well-known risk factor for the development of macro vascular disease. Excess TNF- α can result in harmful inflammatory responses. Whereas too little can contribute to persistent infection. In that regard, some polymorphisms in the TNF- α gene have been associated with susceptibility to certain autoimmune diseases, 32,33 infectious diseases, 4 certain cancers and diabetes mellitus. However, it has been shown to be non-significant in many other inflammatory diseases. Our study also, could not find any association of TNF- α -308 gene polymorphism with diabetic peripheral neuropathy. Similarly, a study in South Indian population also could

not find any association of TNF- α with diabetic peripheral neuropathy. Wery few reports are available on the association of TNF- α gene polymorphisms with risk of diabetic neuropathy. Interestingly, two studies from South Indian population, present study and other study observed a higher frequency of G/A heterozygotes. To reach a more reliable and comprehensive conclusion, a further investigations on functional basis, to elucidate the genetic role of the cytokine responses in the pathogenesis of Diabetic neuropathy is required.

IL-10 gene is an anti-inflammatory cytokine that may regulate the complex network of reactions. The difference in anti-inflammatory profile is determined not only by the levels of IL-10 production with neurological deterioration and also by the functional polymorphisms of the gene. Several functional IL-10 gene polymorphisms have been described. 41 One of the polymorphism that is associated with low, medium or high production of IL-10 situates in the promoter region of the gene at position -1082.⁴² Based on clinical evaluation of diabetic neuropathy, the 'high-producer' IL-10-1082 G/G genotype may be responsible for the down regulation of immune response leading to inflammation in diabetic neuropathy patients. The most prominent feature from the present study is the significant higher frequency of IL-10 (-1082) GG genotype with higher frequency of GG allele among diabetic neuropathic patients. These findings are in agreement with other study. 40 Studies conducted in France and Spain did not confirm any significant association of DM with different genotypes of IL-10 promoter polymorphisms in Caucasians. 43-45 Whereas few reports from Taiwanese, 46 Turkish 47 and Egyptian subjects⁴⁸ has shown association of IL-10-1082 homozygous GG with DM cases.

The SNP at position +874 A/T plays a fundamental role in the induction of IFN γ production. ⁴⁹ In the first intron of the IFN γ gene, there is a CA repeat polymorphism that affects transcription. Moreover, an adenine (A) to thimine (T) transition at position þ874 (intron 1) has been associated with increased IFN γ expression. ⁵⁰ The T allele of IFN γ +874A/T provides a binding site for the transcription factor NF-kB, which is able to regulate IFN γ expression. ⁵⁰ It is possible that low IFN γ production will facilitate not as much of immune response against inflammation rendering

these individuals more susceptible to the disease as the downstream process would eventually leads to nerve damage. Which is significantly associated with diabetic neuropathy. The same trend was observed in another South Indian diabetic neuropathy cases. Egyptian diabetic cases and Greece diabetic populations and a North Indian population with cervical cancer. See the disease as the disease as the downstream process as the downstream process as the downstream process as the downstream process. All process as the downstream process

The pathogenetic vision of diabetes mellitus has changed in the last several years, with inflammatory pathways playing major roles in the development and progression of diabetic complications. Many of the proposed mechanisms are interdependent and it is likely that more than one mechanism is involved in the development of the chronic complications of diabetes. Understanding the role of risk factors and molecular biomarkers may help in early diagnosis and risk prediction of the condition. In addition to that, there may be genetic influences in either protecting or making them susceptible to the development of complications. Effective treatment and/or prevention will therefore require an integrated approach combining multiple strategies to target the underlying inflammatory processes. Diabetic Neuropathy designed to target specific cytokines, chemokines, growth factors and even transcription factors is already well underway. Besides the assessment of pharmacological safety, bioavailability, and efficacy in vivo, more clinical studies will further support the potential of such strategies to be used in diabetes therapy. It is possible that in the coming years the hope of new therapeutic and preventive strategies based on inflammatory properties with beneficial actions on diabetic complications can be translated into real clinical treatment.

CONCLUSIONS

In conclusion, this case-control study suggests that IL-10polymorphism is associated with the susceptibility to diabetic neuropathy in type 2 DM patients. The results shows an increased risk, indicating that individuals with GG phenotypes of IL-10 and AA phenotypes of IFNy are two times more likely to get the disease. Analysis of gene-gene interaction of the 3 factors that may affect the diabetic neuropathy shows that there was an interaction between the three SNP's. IL-10 serves as an important biomarker in Indian population for their susceptibility to Diabetic Neuropathy as it may play a role in alteration of IL-10 production and the inflammatory responses during the course of the disease. Repeated studies with large sample size are required for further validation of our findings and also to understand the association between cytokine gene polymorphisms and the development of Diabetic Neuropathy.

ACKNOWLEDGEMENTS

The authors wish to thank Prof. Jason H. Moore - Director, and Peter Andrews - Programmer, Institute for Quantitative Biomedical Sciences, Dartmouth-Hitchcock

Medical Center, Lebanon, for their helpful comments and technical assistance given at the time of multifactor dimensionality reduction [MDR] analysis. In addition, the authors would also thank the patients and healthy subjects, who willingly participated in the study.

Funding: No funding sources Conflict of interest: None declared Ethical approval: The study was approved by the local (Andhra University) ethics committee

REFERENCES

- Christian N. Diabetes Cases Rise From 30 Million to 230 Million in 20 Years. In: Medical News Today (ed). International Diabetes Federation; 2006 (available online http://www.medicalnewstoday. com/articles/44967.php). (Retrieved on 11/6/2006).
- Daneshmandi S, Pourfathollah A, Arababadi MK, Hassanshahi G, Rezaeian M, Asiabanha M. Evaluation of relation between IL-4 and IFN- γ polymorphisms and type 2 diabetes. Mazand Univ Med Sci.2008; 18:35-41.
- 3. Reutens AT, Atkins RC. Epidemiology of diabetic nephropathy. Contrib Nephrol. 2011; 170: 1–7.
- 4. Wong MC, Chung WY, Wong KS. Effects of treatments for symptoms of painful diabetic neuropathy: systematic review. BMJ. 2007; (2): 1-10.
- 5. Braunwald E, Fauci AS, Kasper DL. In Harrison Principles of Internal Medicine McGraw- Hill (ed), 2001; vol 2, 15th edition. New York.
- 6. Feldman EL, Russell JW, Sullivan KA, Golovoy D. New insights into the pathogenesis of diabetic neuropathy. Curr Opin Neurol. 1999; 12: 553-563.
- 7. Quasthoff S. The role of axonal ion conductances in diabetic neuropathy: a review. Muscle Nerve. 1998; 21: 1246-1255.
- 8. Martinez-Blasco A, Bosch-Morell F, Trenor C, Romero FJ. Experimental diabetic neuropathy: role of oxidative stress and mechanisms involved. Biofactors 1998; 8: 41-43.
- World Health Organization. Diabetes: The Cost of Diabetes Fact Sheet #236 2002(available online http://www.who.int/mediacentre/factsheets/fs236/en/). (Retrieved on 12/8/2005)
- Kovar and Florian M. In The Tumor Necrosis Factor [Alpha] -308 G/A Polymorphism Does Not Influence Inflammation And Coagulation Response In Human Endotoxemia. 2007; pp 238-241. Vol 27 -Issue 3.
- 11. Hotamisligil GS, Arner P, Caro JF, Atkinson RL, Spiegelman BM. Increased adipose tissue expression of tumor necrosis factor-alpha in human obesity and insulin resistance. J. Clin. Invest. 1995; 95, 2409–2415.
- Wastowski IJ, Peres NT, Simões RT, Castelli EC, Simões AL, Martinez-Rossi NM, Donadi EA. Identification of a novel 120 bp allele at the TNFd microsatellite locus. Tissue Antigens. 2006; 67(4):318–20.

- 13. Hajeer AH, Hutchinson. Influence of TNF alpha gene polymorphisms on TNF alpha production and disease. Hum Immunol. 2001; 62: 1191–1199.
- Dokter WH, Koopmans SB, Vellenga E. Effects of IL-10 and IL-4 on LPS-induced transcription factors (AP-1, NF-IL-6, NFκB) which are involved in IL-6 regulation. Leukemia. 1996; 10: 1308.
- 15. Korholz D, Banning U, Bonig H, et al. The role of Interleukin-10 in IL-15-mediated T cell responses. Blood. 1997; 90: 4513.
- 16. Stalenhoef JE, Alisjahbana B, Nelwan EJ, et al. The role of interferon-gamma in the increased tuberculosis risk in type 2 diabetes mellitus. Eur J Clin Microbiol Infect Dis. 2008; 27:97-103.
- 17. Cantor MJ, Nickerson P, Bernstein CN. The role of cytokine gene polymorphisms in determining disease susceptibility and phenotype in inflammatory bowel disease. Am J Gastroenterol. 2005; 100:1134–42.
- 18. Lahari DK, Bill S. DNA isolation by a rapid method from human blood samples: Effects of MgCl2, EDTA, Storage Time and Temperature on DNA yield and quality. Biochem. Genet. 1993; 31 (7&8):321-328.
- Perrey C, Turner SJ, Pravica V, Howell WM, Hutchinson IV. ARMS-PCR methodologies to determine IL-10, TNF-alpha, TNF-beta and TGFbeta1 gene polymorphisms. Transpl Immunol. 1999; 7:127–8.
- Balakrishnan V. Hardy-weinberg equilibrium and allele frequency estimation. In: Malhotra KC (ed) Statistical Methods in human population genetics, IBRAD, ISI and ISHG, Calcutta. 1988; pp. 39-93.
- 21. Taylor GL, Prior AM. Blood groups in England II distribution in the population. Ann Eugen. 1938; 8: 356 361.
- 22. DerSimonian R, Laird N. Meta-analysis in clinical trials. Control Clin Trials. 1986; 7: 77–88.
- 23. Ritchie MD, Hahn LW, Moore JH. Power of multifactor-dimensionality reduction for detecting gene-gene in teractions in the presence of genotyping error, missing data, phenocopy, and genetic heterogeneity. Genet Epidemiol. 2003; 24, 150–157.
- 24. Moore JH. Computational analysis of gene-gene interactions using multifactor dimensionality reduction. Expert Rev Mol Diagn 4, 795–803.
- 25. Tesfaye S, Stevens LK, Stephenson JM, Fuller JH, Plater M, Ionescu-Tirgoviste C, Nuber A, Pozza G, Ward JD (1996) Prevalence of diabetic peripheral neuropathy and its relation to glycemic control and potential risk factors. The Euro Diab IDDM complications study. Diabetologia. 2004; 39: 1377-1384
- 26. Shiba T, Higashi N, Nishimura Y. Hyperglycemia due to insulin resistance caused by interferongamma. Diabet Med. 1998; 15:435-6.
- 27. Yu LN, Yang XS, Hua Z and Xie W. Serum levels of pro-inflammatory cytokines in diabetic patients with peripheral neuropathic pain and the correlation

- among them. Zhonghua Yi Xue Za Zhi. 2009; 89: 469-471.
- 28. Gonzalez JFN and Fernandez CM. The role of inflammatory cytokines in diabetic nephropathy. J. Am. Soc. Nephrol. 2008; 19: 433-442.
- Doupis J, Lyons TE, Wu S, Gnardellis C, Dinh T and Veves A. Microvascular reactivity and inflammatory cytokines in painful and painless peripheral diabetic neuropathy. J. Clin. Endocrinol. Metab. 2009; 94: 2157-2163.
- 30. Andreakos ET, Foxwell BM, Brennan FM, Maini RN, Feldmann M. Cytokines and anti-cytokine biologicals in autoimmunity: present and future. Cytokine Growth Factor Rev. 2002; 13:299–313.
- 31. Pfeffer K. Biological functions of tumor necrosis factor cytokines and their receptors. Cytokine Growth Factor Rev. 2003; 14:185–91.
- 32. Brinkman BM, Huizinga TW, Kurba SS et al. Tumour necrosis factor a gene polymorphisms in rheumatoid arthritis: association with sus ceptibility to, or severity of, disease? Br J Rheumatol. 1997; 36:516–21.
- 33. Rood MJ, van Krugten MV, Zanelli E et al. TNF-308A and HLADR3 alleles contribute independently to susceptibility to systemic lupus er ythematosus. Arthritis Rheum. 2000; 43:129–34.
- 34. Yoon JK, Hyo-Suk L, Jung-Hwan Y et al. Association of TNF-a pro moter polymorphisms with the clearance of hepatitis B virus infection. Hum Molec Genet. 2003; 12:2541–6.
- 35. Juszczynski P, Kalinka E, Bienvenu J et al. Human leukocyte antigens class II and tumor necrosis factor genetic polymorphisms are inde pendent predictors of non-Hodgkin lymphoma outcome. Blood. 2002; 100:3037–40.
- 36. Kubaszek A, Pihlajamaki J, Komarovski V et al. Promoter polymorphisms of the TNF-α (G-308A) and IL-6 (C-174G) genes predict the conversion from impaired glucose tolerance to type 2 diabetes. Diabetes. 2003; 52: 1872–1876.
- 37. Mascher B, Schmitt W, Csernok E, Tatsis E, Reil A, Gross WL. Polymorphisms in the tumor necrosis factor genes in Wegener's granulomatosis. Exp Clin Immunogenet. 1997; 14:226–33.
- Gencik M, Borgmann S, Zahn R, Albert E, Sitter T, Epplen JT. Immunogenetic risk factors for antinetrophil cytoplasmic antibody (ANCA) associated systemic vasculitis. Clin Exp Immunol. 1999; 117:412.
- 39. Shiau MY, Wu CY, Huang CN, Hu S-W, Lin SJ, Chang YH. TNF-polymorphisms and type 2 diabetes mellitus in Taiwanese patients. Tissue Antigens. 2003; 61:393–7.
- 40. Kolla VK, Madhavi G et al. Association of tumor necrosis factor alpha, interferon gamma and interleukin 10 gene polymorphisms with peripheral neuropathy in South Indian patients with type 2 diabetes. Cytokine. 2009; 47 (3): 173-7.
- 41. Koch W, Kastrati A, Bottiger C, Mehilli J, von Beckerath N, Schomig A. Interleukin-10 and tumor

- necrosis factor gene polymorphisms and risk of coronary artery disease and myocardial infarction. Atherosclerosis. 2001; 159, 137–144.
- 42. Clerici M, Merola M, Ferrario E, Trabattoni D, Villa ML, Stefanon B et al. Cytokine production patterns in cervical intraepithelial neoplasia: Association with human papillomavirus infection. J Natl Cancer Inst. 1997; 89:245–50.
- 43. Urcelay E, Santiago JL, de la Calle H, Martinez A, Figueredo A, Fernandez-Arquero M. et al. Interleukin-10 polymorphisms in Spanish type 1 diabetes patients. Genes. Immun. 2004; 5, 306–309.
- 44. Reynier F, Cazalis MA, Lecoq A, Paye M, Rosa A, Durand A et al. Lack of association of IL-10 promoter gene variants with type 1 diabetes in a French population. Hum. Immunol. 2006; 67, 311–317.
- 45. Scarpelli D, Cardellini M, Andreozzi F et al. Variants of the interleukin 10 promoter gene are associated with obesity and insulin resistance but not type 2 diabetes in Caucasian Italian subjects. Diabetes. 2006; 55(5): 1529-33.
- 46. Kung WJ, Lin CC, Liu SH, Chaung HC. Association of interleukin-10 polymorphisms with cytokines in type 2 diabetic nephropathy. Diabetes Technol Ther. 2010; 12(10):809-13.
- 47. Hassan Mohebbatikaljahi, Sevda Menevse1, Ilhan Yetkin and Huseyin Demirci. Study of interleukin-10 promoter region polymorphisms (-1082A/G, -819T/C and -592A/C) in type 1 diabetes mellitus in Turkish population. Journal of Genetics. 2009; Vol. 88, No. 2.
- 48. Helaly MAH, El-Sayed ZH, Megahed AE, Ibrahem EF, Afaf Alsaid, Ibrahim AA, Ahmad Settin.

- Association of IL-10 and IL-6 Gene Polymorphisms with Type 2 Diabetes Mellitus among Egyptian Patients. Eur J Gen Med. 2013; 10(3):158-162
- Vandana AG, Carrara Henri RO, Sachs Johnny A, Hoffman Margaret, Stanczuk Grazyna A, Williamson Anna-Lise. Ethnic differences in allelic distribution of IFN-gamma in South African women but no link with cervical cancer. J Carcinogen. 2003; 2:1–8.
- 50. Pravica V, Perrey C, Stevens A, Lee JH, Hutchinson IV. A single nucleotide polymorphism in the first intron of the human IFN-gamma gene: absolute correlation with a polymorphic CA microsatellite marker of high IFN-gamma production. Hum Immunol. 2000; 61: 863–6.
- 51. Tsiavou A, Hatziagelaki E, Chaidaroglou A, Koniavitou K, Degiannis D, Raptis SA. Correlation between intracellular interferon-gamma (IFN-gamma) production by CD4+ and CD8+ lymphocytes and IFN-gamma gene polymorphism in patients with type 2 diabetes mellitus and latent autoimmune diabetes of adults (LADA). Cytokine. 2005; 31(2): 135-41.
- 52. Tamandani MKK, Sobti RC, Shekari M, Mukesh M, Suri V. Expression and polimorphism of IFN-gamma gene in patients with cervical cancer. Exp Oncol. 2008; 30:224-9.

DOI: 10.5455/2320-6012.ijrms20141142 **Cite this article as:** Ramesh M, Kumari KG, Sudhakar G. The cytokine gene polymorphisms (TNF- α , IL-10 And IFN- γ) and the role of inflammatory cytokines in diabetic neuropathy. Int J Res Med Sci 2014:2:1470-8.