

Research Article

Microbiological study of neonatal sepsis in a tertiary care hospital of western Uttar Pradesh, India

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ABSTRACT

Background: Neonatal sepsis is a clinical syndrome characterized by systemic signs of infection and accompanied by bacteraemia in the first month of life. Neonatal sepsis is among the three most common illnesses among newborns, and second most common cause of neonatal mortality especially among preterm and low birth weight babies. Aim of the study was to identify the common bacterial pathogens associated with neonatal sepsis and their antibiotic susceptibility pattern.

Methods: Total 120 blood samples collected aseptically (from January 2015 to April 2015) and cultured in automated machine Versa TREK™ automated microbial detection system. Significant bacterial colonies are seen on streak lines which were observed by their diameter, opaqueness, flatness, regular or irregular margins etc. Any growth was subjected for identification by appropriate biochemical tests. Antibiotic susceptibility testing was done.

Results: Out of one hundred twenty (120) samples, eighty (66.67%) samples were found positive for bacterial infection. In which 22 (27.50%) *E.coli*, 14 (17.50%) *Klebsiella pneumoniae*, 10 (12.50%) *pseudomonas*, 17 (21.25%) *Staphylococcus aureus*, 6 (7.50%) coagulase negative staphylococci (CoNS) and 11(13.75%) *Enterococcus sp.* was identified.

Conclusions: The blood culture positivity rate was 66.67%. *E. coli*, *Klebsiella spp.*, and *Staphylococcus aureus* was the commonest agent.

Keywords: Neonatal septicaemia, Blood culture, Antibiotic susceptibility pattern

INTRODUCTION

Neonatal sepsis is a clinical syndrome characterized by systemic signs of infection and accompanied by bacteraemia in the first month of life. Neonatal sepsis is among the three most common illnesses among newborns, and second most common cause of neonatal mortality especially among preterm and low birth weight babies.¹

According to world health organisation estimate, there are about 5 million neonatal deaths each year, 98% of death occurring in develop countries. Neonatal sepsis is responsible for 30% to 50% of total neonatal deaths each year in developing countries. Infections are estimated to

cause 1.6 million deaths annually. Sepsis and meningitis are the most commonly implicated factor for most of these death.² In India, the incidence of blood culture proven sepsis was reported as 8.5 per 1,000 live births for the year 2002–2003 by the National Neonatal Prenatal Database.³

Most of the neonatal sepsis related deaths are preventable if suspected early and treated with appropriate antibiotics. Neonatal sepsis is broadly categorized into early and late onset sepsis depending upon the postnatal day of presentation. Early-onset neonatal sepsis (EONS) occurs within first 72 h of life, while the late-onset neonatal sepsis (LONS) occurs between 72 h to 90 days of life.^{4,5} Because of the different pathogens involved in neonatal

sepsis of onset, appropriate management and care depends on knowledge about the causative organisms and their sensitivity to antibiotics. The present study focused on isolation and determination of antimicrobial activity of neonatal sepsis causing bacteria.

METHODS

Study was conducted in NICU, Dept. of Microbiology, LLRM Medical College, Meerut, Uttar Pradesh, India. Total 120 blood samples collected from January 2015 to April 2015 aseptically and cultured in automated machine Versa TREK™ automated Microbial Detection system. After the incubation of 5-6 hours positive samples sub cultured on blood agar and MacKoncy agar and incubate at 37°C for overnight. Significant bacterial colonies are seen on streak lines which were observed by their diameter, opaqueness, flatness, regular or irregular margins etc. Identification is done by biochemical test, drug susceptibility test also done by Kirby bauer disc diffusion method.

RESULTS

Out of one hundred twenty (120) samples, eighty (66.67%) samples were found positive for bacterial infection. Gram negative isolates (57.5%) were more as

compared with Gram positive isolates (42.5%). In which 22 (27.50%) *E.coli*, 14 (17.50%) *Klebsiella pneumonia*, 10(12.50%) *pseudomonas*, 17 (21.25%) *Staphylococcus aureus*, 6 (7.50%) coagulase negative *staphylococci* (CoNS) and 11 (13.75%) *Enterococcus sp.* was identified (Table 1).

Table 1: Isolated bacterial species and their percentage from blood sample.

Isolated bacteria	No. of cases	Percentage
<i>E. coli</i>	22	27.50%
<i>Klebsiella species</i>	14	17.50%
<i>Pseudomonas aeruginosa</i>	10	12.50%
<i>Staphylococcus aureus</i>	17	21.25%
CoNS	6	7.50%
<i>Enterococcus</i>	11	13.75%
Total	80	100%

E.coli was 100% resistant to aztreonam and gentamycin, *Klebsilla* was 100% resistant to imipenem, Aztreonam and gentamycin, *Pseudomas aerugnosa* was 100% resistant to imipenem, Polymixin-B, Pipracillin/Tazobactam, Aztreonam and gentamycin (Table 2).

Table 2: Antibiotics resistance pattern shown by gram negative bacterial isolates from blood sample.

Gram negative organism antibiotics	Number and percentage of resistance cases		
	<i>E. coli</i> (n=22)	<i>Klebsiella</i> (n=14)	<i>Pseudomonas</i> (n=11)
Meropenem (ME)	16 (72.7)	8 (57.1)	7 (63.6)
Imepenem (IM)	16 (72.7)	14 (100)	11 (100)
Colistin (CL)	11 (50)	9 (64.2)	2 (18.1)
Polymixin-B (PB)	18 (81.8)	10 (71.4)	11 (100)
Piperacillin/Tazobactam (Pt)	19 (86.3)	13 (92.8)	11 (100)
Amikacin (AK)	20 (90.9)	12 (85.7)	1 (9.09)
Netilmicin (Nt)	20 (90.9)	13 (92.8)	3 (27.2)
Azteronam (AT)	22 (100)	14 (100)	11 (100)
Gentamycin (G)	22 (100)	14 (100)	11 (100)

Table 3: Antibiotics resistance pattern shown by gram positive bacterial isolate from blood sample.

Gram positive organism antibiotics	Number and percentage of resistance cases		
	<i>S.aureus</i> (n=17)	CoNS (n=6)	<i>Enterococci</i> (n=11)
Vancomycin (VA)	10 (58.8)	3 (50)	8 (72.7)
Teicoplanin (TEI)	9 (52.9)	3 (50)	8 (72.7)
Linezolid (LZ)	9 (52.9)	3 (50)	5 (45.4)
Amikacin (AK)	17 (100)	6 (100)	11 (100)
Netlimicin (Nt)	17 (100)	6 (100)	11 (100)
Cefoxitin (CX)	17 (100)	6 (100)	11 (100)
Erythromycin (ERT)	17 (100)	6 (100)	11 (100)
Gentamycin (G)	17 (100)	6 (100)	11 (100)
Polymixin-B (PB)	17 (100)	1 (100)	11 (100)

S.aureous was 100% resistant to Amikacin, Netilmicin, Cefoxitin, Erythromycin, Gentamycin and Polymixin-B, CoNS was 100% resistant to Amikacin, Netilmicin, Cefoxitin, Erythromycin, Gentamycin, *Enterococci* was 100% resistant to Amikacin, Netilmicin, Cefoxitin, Erythromycin, gentamycin and Polymixim-B (Table 3).

DISCUSSION

Neonatal septicaemia is one of the major causes of mortality and morbidity. Septicaemia usually consists of bacteraemia plus a constellation of signs and symptoms caused by the microorganisms or their toxic products in the circulation. The usual source of infection include incubators, resuscitators, ventilators, solution for could sterilization, feeding bottles, catheters, face masks, and infusion set and sites etc.⁶ Sepsis is a significance cause of death in the newborn, particularly among those of very low birth weight and premature infants.

The world health Organization (WHO) estimate that worldwide 1.6 million newborn baby die every year from neonatal infections. Despite recent advances in neonatal intensive care and current strategies to treat neonatal sepsis, mortality rates have not fallen for over three decades except in babies born to mothers who have received intrapatum prophylaxis (IAP) for group B streptococcus.⁷

The total blood culture positivity rate among neonates with sepsis in this study group was 66.67%, which was comparable to the study by Higher culture positivity rates of 25.6%, 28.3%, 48%, 62.8% and 64% among neonates with sepsis had been reported. In this study the gram negative rods were the main cause of septicaemia.⁸⁻¹²

Identified as *E.coli* 22 (27.50%), *Klebsiella pneumonia* 14(17.50%), *Pseudomonas* 10 (12.50%), this is approximately same as Rao Pooja et al says that *Klebsiella spp.* (15.5%), in another study isolation rate of *K. pneumonia*, *S. aureus*, *E. coli* were 22.41%, 18.37%, 6.12% respectively.^{6,20} Emergence of antimicrobial resistance in GNB poses major therapeutic challenge. Many studies have shown resistance to third-generation cephalosporins, aminoglycosides, and carbapenems in recent years.^{13-19,21}

In the present study, GNB were highly resistance to Azteronam (AT) (100%), Gentamycin (G) (100%).The most common causes of death in the neonatal period are infections, including septicaemia, meningitis, respiratory infections, diarrhoea, and neonatal tetanus (32%), followed by birth asphyxia and injuries (29%), and prematurity (24%).

The data available are a mixture of official sources and hospital and community based studies. In developing countries, the rate of home deliveries is high, and the percentage of deliveries assisted by a skilled attendant is low: in Africa it ranges from 37% in sub-Saharan Africa

to 69% in North Africa, in Asia from 29% in South Asia to 66% in East Asia and the Pacific region. In South America and the Caribbean, it is about 83%.²

CONCLUSION

To conclude, in our study, the blood culture positivity rate was 66.67%. Gram negative organisms were predominant. *E.coli* 22 (27.50%), was the most common isolate.

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Ethical approval: The study was approved by the Institutional Ethics Committee

REFERENCES

1. Morris A. Neonatal Septicaemia. *Pediatr. Clin. N. Amer.* 1983;30:2.
2. Vergnano S, Sharland M, Kazembe P, Mwansambo C, Heath PT. Neonatal sepsis: an international perspective; *Arch Dis Child Fetal Neonatal.* 2005;90:F220-4.
3. National Neonatal Perinatal Database. [Internet]. NNPD report 2002-03. Available from: http://www.newbornwhocorg/pdf/nnpd_report_2002-03.pdf; 2005.
4. Sundaram V, Kumar P, Dutta S, Mukhopadhyay K, Ray P, Gautam V, et al. Blood culture confirmed bacterial sepsis in neonates in a North Indian tertiary care center: changes over the last decade. *Jpn J Infect Dis.* 2009;62:46-50.
5. Anderson-Berry AL, Bellig LL, Ohning BL. Neonatal sepsis. *emedicine Pediatrics: Cardiac disease and critical care medicine.* 2010; 978352 [Updated 2010 Feb 23; Cited 2010 Sep 22].
6. Cassone A, Bernardis De, Mondello F, Ceddia T, Agatensi L. Evidence for a correlation between mortality and morbidity in neonates. *J. Infect. Dis.* 2003;156:777-83.
7. Al-Sshamahy HA, sabrah AA, Al-robasi BA, Naser MS.. Types of bacteria associated with neonatal sepsis in Al- Thawra University Hospital, Sana'a, Yemen, and their antimicrobial profile. *SQU Med.* 2012;12:48-54.
8. Awaisua A, Sulaiman SA, Ibrahim MI, Saad A. Antimicrobials utilization and outcomes of neonatal sepsis among patients admitted to a University teaching hospital in Malaysia. *Eastern J Med.* 2007;12:6-14.
9. Jain NK, Jain VM, Maheshwari S. Clinical profile of neonatal sepsis. *Kathmandu Univ Med J (KUMJ)* 2003;1:117-20.
10. Bhattacharjee A, Sen MR, Prakash P, Gaur A, Anuprba S. Increased prevalence of extended spectrum beta lactamase producers in neonatal septicaemic cases at a tertiary referral hospital. *Indian J Med Microbiol.* 2008;26:356-60.

11. Rahman S, Hameed A, Roghani MT, Ullah Z. Multidrug resistant neonatal sepsis in Peshawar, Pakistan. *Arch Dis Child Fetal Neonatal.* 2002;87:F52-4.
12. Tallur SS, Kasturi AV, Nadgir SD, Krishna BV. Clinico-bacteriological study of neonatal septicemia in Hubli. *Indian J Pediatr.* 2000;67:169-74.
13. Report of the national neonatal perinatal database national neonatology Forum; 2002-2003.
14. Bang AT, Bang RA, Baitule S, Deshmukh M, Reddy MH. Burden of morbidities and the unmet need for health care in rural neonates – a prospective observational study in Gadchiroli, India. *Indian Pediatr.* 2001;38:952-65.
15. Hossain MM, Afroza S, Shirin M, Chowdhury NA, Saha SK. Bacterial aetiology of neonatal sepsis in a tertiary care hospital in Bangladesh. *Bangladesh J Child Health.* 2004;28:81-5.
16. Kumar M, Paul VK, Kapoor SK, Anand K, Deoraria AK. Neonatal outcomes at a subdistrict hospital in North India. *J Trop Pediatr* 2002; 48:43-6.
17. Saxena S, Anand NK, Saini L, Mittal SK. Bacterial infections among home delivered neonates. Clinical picture and bacteriological profile. *Indian Pediatr.* 1980; 17:17-24.
18. Mathur NB, Singh A, Sharma VK, Satyanarayana L. Evaluation of risk factors for fatal neonatal sepsis. *Indian Pediatr.* 1996;33:817-22.
19. Kapoor L, Randhawa VS, Deb M. Microbiological profile of neonatal septicemia in a pediatric care hospital in Delhi. *J Commun Dis.* 2005;37:227-32.
20. Rao P, Sowmya KN, Shrikala B, Radhakrishna M, Keerthiraj B. A Spectrum of Bacterial Pathogens and its Antibiotic Susceptibility Pattern Isolated from Neonatal Sepsis in an NICU in a Government Pediatric Hospital; *Research Journal of Biological Sciences.* 2015;4(5):50-54.
21. Sheth KV, Patel TK, Tripathi CB. Antibiotic Sensitivity Pattern In Neonatal Intensive Care Unit of A Tertiary Care Hospital of India; *Asian Journal of Pharmaceutical and Clinical Research.* 2012;5(3).

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